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Ring opening of 2-phenylazetidines with allylsilanes

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Abstract—N-Activated 2-phenylazetidines were opened regioselectively at the benzylic carbon with various allylsilanes or propargylsilane in the presence of BF_3Et_2O , providing amino olefins, precursors of biomolecules such as phenyl-homo-kainoids. 2006 Elsevier Ltd. All rights reserved.

Over the years the reactivity and/or use of aziridines and azetidines have been addressed by several groups. $1-13$ Shortly we and others have reported on new reactivities of N-activated 2-phenylaziridines and 2-phenylazetidines towards various nucleophiles. These three and four membered heterocycles have been identified, respectively, as masked 1,3 and 1,4 dipoles in the presence of selected dipolarophiles.[14–18](#page-2-0) Recently we focused on the reactivity of 2-phenylazetidines (1a–b) as electrophiles towards various allylsilanes, and report here our preliminary results on the successful ring opening of the azetidines 1a–b together with relevant applications towards the preparation of biomolecules.

The two starting azetidines 1a (NTs) and 1b (NNos), selected for our study can be readily obtained from styrene following reported procedures.^{[19](#page-2-0)} The reaction of $1a$ or 1b with trimethylallylsilane 2 was performed at room temperature in CH_2Cl_2 in the presence of $BF_3 OEt_2$ ([Scheme 1](#page-1-0)). After 30 min reaction time, no starting azetidine was apparent on TLC. From the reaction mixture two adducts could be isolated by column chromatography, and identified as $3a-a'$, a mixture of inseparable silylated piperidines, and 4a the open chain allylated adduct in a ratio $3a-a'/4a$: 1/4. This first experiment established that azetidine 1a is ring opened with allylsilane at the benzylic carbon via a $S_E\bar{2}$ pathway where azetidine 1a is the electrophilic substrate (Eq. 1). The piperidine adducts $3a-a'$ is resulting from an

internal collapse of the silicon stabilized b-carbenium on the transient amide anion. A similar result was observed with the corresponding aziridine.^{[7](#page-2-0)} It is noteworthy that the silylated piperidines $3a-a'$ could be transformed quantitatively into 4a via a brief heating in a TBAF solution. Interestingly if the initial reaction mixture was aged overnight only the allylated adduct 4a was isolated, in situ desilylation being promoted by the fluorinated Lewis acid. This observation will simplify the purification step, because at that time the allylated adducts were our targets for further applications (vide infra). Very similar results in reaction rates and yields were obtained with 1b, the nosyl activated azetidine, in this case we isolated the adducts $3b-b'$ and $4b$, respectively, the piperidine and the allylated adducts. The mixture of piperidines $3b-b'$ is cleanly converted to 4b by a TBAF treatment or more conveniently with the overnight ageing procedure. As anticipated the tosyl or nosyl activations in 1a or 1b are similarly effective for the azetidine ring opening with allylsilane 2. Conversely when different Lewis acids (TiCl₄, $MgBr_2$, $ZnBr_2$) were experimented, we observed severe contaminations with byproducts, mainly halogenated adducts at the benzyl carbon. Therefore BF_3E_5O was favoured as a good compromise between electronic assistance and intrinsic reactivity. The temperature seemed not to be a critical factor but we found that the ideal conditions are to mix azetidine (1a) and allylsilane (2) at -78 °C, followed by dropwise addition of the Lewis acid. The mixture was then left to warm up to room temperature overnight. The remarkable regioselectivity in the ring opening at the benzylic carbon is best explained by the release of the cyclic constraint and the polarization of the benzylic $C-N$ bond with the Lewis acid.^{[20](#page-2-0)}

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Scheme 1. Reagents and conditions: (i) BF_3Et_2O , CH_2Cl_2 from -78 °C to rt; (ii) TBAF in THF reflux 1 h.

Next the reactivity of easily accessible allylsilanes namely the E crotylsilane (5), or 2-chloromethyl-3-trimethylsilylpropene (7) towards 1a–b was explored (Eqs. 2 and 3). In each case the expected transformations were operative and delivered the allylated adducts $6a-a'$, $6b-b'$, mixtures of diastereomers ($1/1$ ratio), and $8a$. In a complementary study, we decided to oppose propargylsilane (9) to azetidine 1a and 1b (Eq. 4). Surprisingly even if propargylsilane is known to be less nucleophilic than allylsilane, the allenyl adducts 10a and 10b were isolated in acceptable yields. These data demonstrated that azetidines of type 1a or 1b are of great value as masked electrophiles.

To explain the regioselective ring opening of azetidines 1a–b, with allylsilanes or propargylsilane, we propose the mechanism depicted in Scheme 2. The Lewis acid $(BF_3 \cdot Et_2 O)$ may have a double function (i) first in coordinating the sulfonyl appendage that stretch out the benzylic C–N bond leading to the electrophilic substitution; (ii) second in stabilizing the transient amide in intermediate A, which can then evolve to a mixture of 3a and 4a, or solely to 4a, respectively, if the reaction is quenched after 1 h, or left overnight (Scheme 2). Then we decided to prepare more complex allylsilanes 11, 12, 14 and 16^{21} 16^{21} 16^{21} in order to extend the scope of the reaction and to apply it to the synthesis of scaffolds occurring in biomolecules (Eqs. 5–8). Each reaction with these allylsilanes deserves special comments. With bis-allylsilane 11 only adduct 4a was obtained, the expected cooperativity and/or double addition of the two allylsilane residues embedded in 11 was not detected. Considering the peculiar reactivity of azetidine 1a with methylenecyclohexane,^{[15](#page-2-0)} allysilane 11 is combining two potential nucleophilic centres one on the alkene and one on the allylsilane. Therefore a multi-path reaction was awaited for 11 towards 1a, via two putative carbenium ions on the β - or γ -carbon next to the silicon atom.

Indeed the reaction proceeded, but only adduct 13a could be characterize, accounting that a carbenium stabilized by the silicon atom ranked over a tertiary one. Similarly as in our work with aziridines azetidines 1a and 1b were treated with cyclopentenyl- (14) and cyclohexenyltrimethysilane (16), the allylation worked efficiently, delivering inseparable mixtures (always in $1/1$ ratio) of $15a-a'$, $15b-b'$ and $17a-a'$, respectively, in 65% and 70% yield. Indeed we found a practical and efficient reaction for the regioselective ring opening of azetidines with allylsilanes. As we are currently interested in the preparation of kainoids, 22 we considered that $15a-a'$ may constitute ideal starting materials for the obtention of homo-kainoids ([Scheme 3](#page-2-0)). Kainoids have been used as successful probes to understand the excitatory functions of glutamic acid in the mammalian central nervous system. $23-25$ But there is still a demand for more selective ligands towards the kainate receptors, especially for antagonists, and homo-kainoids may be potential candidates. Indeed following our previous reported sequence for the synthesis of phenyl-kainic acid, we used $15a-a'$ towards the formal synthesis of

Scheme 2. Proposed mechanism for the regioselective opening of azetidine 1a.

Scheme 3. Reagents and conditions: (i) $Pd(OAc)₂, O₂, DMSO, 40 °C$, 4 h, separation via chromatography; (ii) $RuCl₃$, $NaIO₄$; (iii) $CH₂N₂$ in $Et₂O.$

phenyl-homo-kainic acid. Using the hydroamination reaction assisted by $Pd(II)$,²⁶ the mixture of diastereomers (15a-a') was converted into bicyclic adducts 16a and 17a, which could be separated by column chromatography and identified by extensive NMR (COSY and NOESY experiments) to be, respectively, the exo (16a) and endo (17a) adducts with a cis ring junction. Finally, a single crystal X-ray analysis of 17a confirmed our NMR analysis.²⁷ Then each diastereomer could be transformed into the fully protected phenyl-homokainoids 18a–19a in a two step sequence: Sharpless oxidation and diazomethane esterification, awaiting final deprotection.28,29

In conclusion in this work on N-activated azetidines, we have demonstrated that the regioselective ring opening with allysilanes is realized at the benzylic carbon under smooth reaction conditions in a good yielding sequence. Our work is probably giving a new vitality to the azetidine chemistry, indeed some of the described adducts have a real chemical potential. We are currently using the above methodology towards the synthesis of biological active compounds.

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- 28. Typical experimental conditions for the preparation of $4a$. A mixture of azetidine 1a (100 mg, 0.35 mmol) and trimethylallylsilane $2(85 \mu l, 0.53 \text{ mmol}, 1 \text{ equiv})$ in $\text{CH}_2\text{Cl}_2(5 \text{ ml})$ is cooled at -78 °C and a solution of BF_3 Et₂O (100 µl, 0.8 mmol, 2.3 equiv) in CH_2Cl_2 (1 ml) is added dropwise. After 1 h, $H₂O$ (2 ml) was added and the mixture was left overnight at room temperature. More water was added and the organic layer was separated, washed with brine, dried with $Na₂SO₄$ and concentrated in vacuo. The residue was purified by column chromatography on silica gel eluting with heptane/Et₂O 8:2 to yield $4a$ (70%).
- 29. Selected physical data: Compound $4a$ mp $76-78$ °C; IR (KBr) v (cm^{-1}) 3270, 3090, 1320, 1170; ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3, 25 \text{ }^{\circ}\text{C}) \text{ } \delta \text{ } 7.68 \text{ (d, } J \text{ } 8.5 \text{ Hz, } 2\text{H}) \text{ } 7.38-$ 7.12 (m, 5H), 7.11–7.02 (m, 2H), 5.68–5.52 (m, 1H), 4.95 (d, J 6 Hz, 1H), 4.73 (br s, 1H), 2.74–2.85 (m, 2H), 4.86– 4.83 (m, 1H), 2.62–2.58 (m, 1H), 2.42 (s, 3H), 2.29 (dt, J 7.0 and 1.5 Hz, 2H), 1.97–1.60 (m, 2H); 13C NMR (75 Hz, CDCl₃, 25 °C) δ 143.7, 143.4, 137.1, 136.4, 129.8, 128.7,

127.7, 127.2, 126.6, 116.6, 43.1, 41.6, 41.3, 35.6, 21.7; m/z $(FAB-MS)$ $[M+H^+]$ 330.

Compound 13a, waxy solid; IR (KBr) v (cm⁻¹) 3360, 1640, 1600, 1170; ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 7.64 (d, J 8.3 Hz, 2H), 7.34–7.11 (m, 5H), 7.08–6.97 (m, 2H), 5.49 (dd, J 11 and 17.7 Hz, 1H), 5.25 (dd, J 1.5 and 11.3 Hz, 1H), 4.87 (dd, J 1.5 and 18 Hz, 1H), 4.28 (m, 1H), 2.81– 2.68 (m, 1H), 2.64–2.53 (m, 1H), 2.43 (s, 3H), 2.44–2.36 (m, 1H), 2.05–1.87 (m, 1H), 1.85–1.04 (m, 11H); 13C NMR $(75 \text{ Hz}, \text{ CDCl}_3, 25 \text{ }^{\circ}\text{C})$ δ 143.6, 143, 140.8, 137.4, 130, 128.2, 127.5, 126.9, 116.4, 54.2, 42.4, 38.9, 29.6, 26.8, 22.6, 21.9; m/z (FAB-MS) [M+H⁺] 398.

Compound 16a (exo), mp 123-125 °C; IR (KBr) v (cm⁻¹) 3080, 1610, 1500, 1175; ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 7.76 (d, J 8 Hz, 2H), 7.20–7.36 (m, 5H), 7.09 (d, J 7 Hz, 2H), 5.91 (m, 1H), 5.57 (m, 1H), 4.79 (d, J 5.5 Hz, 1H), 3.48 (m, 1H), 3.30 (m, 1H), 2.90 (m, 1H), 2.76 (m, 1H), 2.46 (s, 3H), 2.25 (m, 1H), 1.93 (m, 2H). 13C NMR $(75 \text{ MHz}, \text{CDCl}_3, 25 \text{ }^{\circ}\text{C})$ d 21.6, 23.0, 32, 38.6, 41.0, 62.9, 126.4, 127. 3, 127.5, 128.5, 129.7, 135.3, 137, 143.2, 143.3; m/z (FAB-MS (M+H⁺).

Compound 17a (endo) mp 85-88 °C; IR (KBr) $v \text{ cm}^{-1}$) 3080, 1615, 1520, 1180; ¹H NMR (300 MHz, CDCl₃, 25 -C) d 7.68 (d, J 8.5 Hz, 2H), 7.35 (d, J 8 Hz, 2H), 7.18– 7.31 (m, 3H), 7.08 (d, J 7 Hz, 2H), 5.80 (m, 1H), 5.38 (m, 1H), 5.07 (s, 1H), 3.97 (d, J 13 Hz, 1H), 2.91 (td, J 12 and 3 Hz, 1H), 2.47 (s, 3H), 2.38 (m, 2H), 2.17 (dd, J 165 and 2.5 Hz, 1H), 1.96 (dd, J 16 and 2.5 Hz, 1H), 1.59 (m, 2H); 13C NMR (75 MHz, CDCl₃, 25 °C) d 21.6, 23, 32, 38.6, 41, 62.9, 1216.4, 127.3, 127.5, 128.5, 129.7, 135.3, 137, 143.2, 143.3; m/z (FAB-MS) 354 (M+H).

Compound 18a waxy solid; IR (KBr) v (cm⁻¹) 3015, 2950, 1710, 1580, 1170; ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 7.82–8.73 (m, 2H), 7.51–7.43 (m, 2H), 7.37–7.18 (m, 3H), 7.17–6.99 (m, 2H), 4.61 (d, J 5.5 Hz, 1H), 4.18–4.01 (m, 1H), 3.62 (s, 3H), 3.42 (s, 3H), 3.58–3.39 (m, 1H), 3.18– 2.89 (m, 2H), 2.79–2.58 (m, 1H), 2.45 (s, 3H), 2.39–2.24 $(m, 1H), 2.19-2.09$ $(m, 2H);$ ¹³C NMR (75 MHz, CDCl₃, 25 -C) d 33.1, 33.8, 40.8, 49.4, 50.5, 51.9, 52.2, 59.0, 64.7, 124.3, 124.5, 127.2, 127.4, 128.3, 128.5, 128.8, 140, 140.7, 145.4, 152.8, 170.2, 172.5; m/z (FAB-MS) 446 (M+H⁺). Compound 19a waxy solid; IR (KBr) v (cm⁻¹) 3010, 2980, 1715, 1510, 1175; ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 7.88 (d, J 8.6, 2H), 7.83 (d, J 9.0, 2H), 7.32–7.23 (m, 3H), 7.07 (d, J 6.7, 2H), 5.13 (d, J 4.9 Hz, 1H), 4.02 (d, J 2.6, 1H), 3.71–3.38 (m, 7H), 2.72–2.52 (m, 2H), 2.47 (s, 3H) 2.17–1.99 $(m, 2H), 1.97-1.77$ $(m, 2H);$ ¹³C NMR (75 MHz, CDCl₃, 25 -C) d 28.5, 33.8, 35.2, 39.9, 49.0, 41.9, 43.5, 52.4, 54.1, 126.5, 126.8, 127.1, 127.7, 128.0, 128.5, 128.9, 143.1, 144, 144.6, 155.1, 171.9, 172.5; m/z (FAB-MS) 446 (M+H⁺).